

Table 1. ^{13}C and ^1H NMR data for **1** (CDCl_3 , TMS as int. standard)

Carbon	$^{13}\text{C}^*$	$^1\text{H}^\dagger J$ (Hz)
1	73.9	6.09 <i>dd</i> 6, 1.5
2	201.9	
3	101.1	5.38 <i>dd</i> 6, 6
4	76.7	4.70 <i>dddd</i> 8, 6, 6, 1.5
5	41.4‡	1.96 <i>ddd</i> 13.5, 8, 5.5 2.30 <i>ddd</i> 13.5, 6, ~0
6	84.1§	4.81 <i>ddd</i> 5.5, 4.5, ~0
7	83.8§	4.74 <i>dd</i> 4.5, 4.5
8	40.7‡	1.73 <i>ddd</i> 13.5, 10, 4.5 2.26 <i>dd</i> 13.5, 6
9	79.7	4.52 <i>ddd</i> 10, 6, 6
10	130.0	5.61 <i>dd</i> 13.5, 6
11	130.5	6.32 <i>dd</i> 13.5, 10
12	133.3	6.42 <i>d</i> 10
13	132.2	
14	29.8	2.57 <i>q</i> 7
15	13.4	1.13 <i>t</i> 7

* 50.10 MHz with the aid of INEPT method.

† 200 MHz.

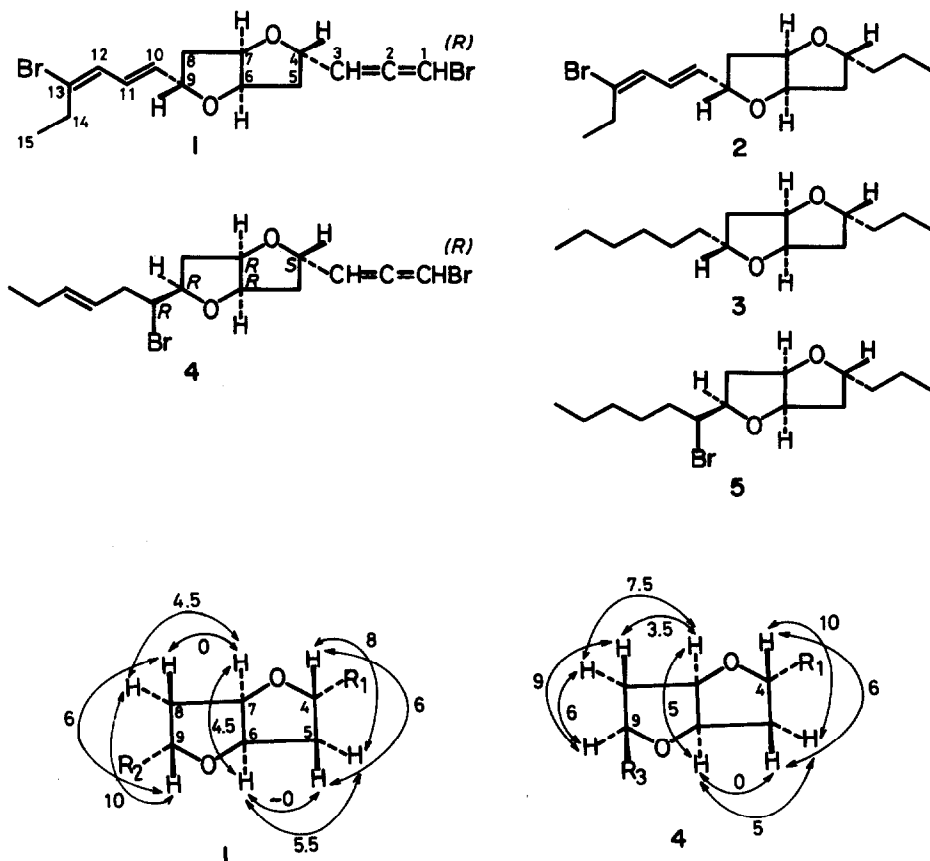
‡, § Assignments may be reversed.

|| Assignments may be interconvertible.

dioxabicyclo[3.3.0]octane skeleton with propyl and C_6 side chains in **2** and **3**.

On detailed analysis of the coupling constants (Fig. 1) in the ^1H NMR spectra of compound **1** and kumausallene (**4**), the stereochemistries at C-4, C-6, C-7 and C-9 were determined. The bromoallenic side chain in kumausallene (**4**) is in the *exo*-configuration in which a pseudo-equatorial conformation can be assumed. As a result the dihedral angle between H_β -5 and H-6 is approximately 90° , corresponding to the observed *J*-value of 0 Hz. However, since the 1-bromo-3-hexenyl side chain in **4** is in the *endo*-configuration, the coupling constant between H-7 and H_β -8 is observed as 3.5 Hz. On the other hand, the coupling constants between H_β -5 and H-6 and between H-7 and H_β -8 in **1** were observed as ~0 Hz and 0 Hz, respectively, and therefore the two side chains at C-4 and C-9 in **1** both had the *exo*-configuration. The *cis* relationship between H-6 and H-7 was also indicated by them having a coupling constant (4.5 Hz) comparable with that of **4**.

Consequently, the relative configurations of the four chiral centres at C-4, C-6, C-7 and C-9 were assigned as in formula **1**. Furthermore, the *E*-configuration of the double bond at C-10 was indicated by the values ($J = 13.5$ Hz in CDCl_3 and $J = 15$ Hz in C_6D_6) of the coupling constants between the pertinent olefinic protons. The double bond at C-12 in **1** was assigned *E*-configuration by the observation of a nuclear Overhauser enhancement (about 10%) between H-11 and H_2 -14.

Fig. 1. The coupling constants (Hz) for the new bromoallene (**1**) and kumausallene (**4**).

In view of the strong negative rotation of **1**, the absolute configuration of the bromoallene moiety in **1** was assigned R-configuration by application of Lowe's rule [14].

Thus, the structure of the new C-15 bromoallene is represented by formula **1**. This is the second example of a C₁₅ non-terpenoid compound from the genus *Laurencia* with a 2,6-dioxabicyclo[3.3.0]octane skeleton.

EXPERIMENTAL

¹H and ¹³C NMR: 100 MHz and 200 MHz, CDCl₃ unless otherwise stated, TMS as int. standard (coupling constant, *J* in Hz); Low and high resolution MS: 70 eV; Optical rotations: CHCl₃; CC: silica gel (Merck, Kieselgel 60, 70–230 mesh); prep. TLC: silica gel 60 F₂₅₄ (Merck). All known metabolites were identified by comparison of the spectral data with those of the authentic specimens. Yields are based on the weights of the extracts.

Collection, extraction and isolation. Samples were collected from six different sites: Kamuimisaki (Shakotan pen., Hokkaido; August 17, 1982), Esashi (Hokkaido; July 24, 1982), Moura (Aomori prefecture; July 23, 1982), Noo (Niigata prefecture; July 6, 1983), Iwaizaki (Shima pen., Mie prefecture; June 30, 1983) and Zaga-shima (Ago Bay, Mie prefecture; June 30, 1983). Extraction and isolation were carried out by conventional methods as described in the case of Zaga-shima's specimen.

Zaga-shima specimen. Half-dried alga (40 g) was extracted with MeOH, and the resulting MeOH soln was treated in the usual manner [1] to give a neutral oil (0.9 g) which was successively fractionated by CC over silica gel. The fraction eluted with hexane was rechromatographed on a silica gel column to yield isolaurene (0.3%) [15], neolaurencenyne (0.5%) [6] and (–)-α-bromocuparene (0.6%)* [16].

The first C₆H₆ fraction was further subjected to prep. TLC to give isolaurinterol (2%) [1] and debromoisolaurinterol (1%) [1, 3]. The next C₆H₆ fraction gave a mixture of laurinterol (17%) [1] and debromolaurinterol (4%) [1], which was separated via their acetates. The last C₆H₆ fraction was repeatedly subjected to a combination of silica gel CC and TLC to yield the new bromoallene **1** (4%) along with prepacifenol (3%) [17], prepacifenol epoxide (1%) [18], pacifenol (9%) [19] and johnstonol (1%) [20]. *Iwaizaki's specimen* contained neolaurencenyne (1.5%) [6], laurencenyne (0.5%) [6], laurinterol (1%) [1], deoxyrepacifenol (0.3%) [21], prepacifenol epoxide (13%) [18], pacifenol (20%) [19] and johnstonol (5%) [20]. *Kamuimisaki, Esashi, Moura and Noo specimens.* These contained laurinterol [1] and debromolaurinterol [1] in 30–40% yields together with neolaurencenyne (ca 1%) [6] and previously reported minor metabolites [3], e.g. isolaurene (ca 1%), (–)-α-bromocuparene (2–3%) and isolaurinterol (3–5%). Laurequinone [7] was also obtained from Moura's and Noo's specimens in 0.3% and 1% yields, respectively.

Compound 1. Oil; $[\alpha]_D^{23} - 215^\circ$ (c 0.985); UV $\lambda_{\max}^{\text{EtOH}}$ nm (ε): 237 (27 000) and 243 (28 000), and $\lambda_{\text{inf}}^{254}$ (19 000); IR ν_{\max}^{film} cm^{–1}: 3060, 1965, 1650, 1610, 1320, 1200, 1190, 1150, 1075, 1030, 990, 965, 915, 885 and 845; ¹H NMR (200 MHz, C₆D₆): δ 0.96 (3H, t, *J* = 7 Hz), 1.24 (1H, ddd, *J* = 13.5, 10, 4.5 Hz), 1.50 (1H, ddd, *J* = 13.5, 8, 5.5 Hz), 1.95 (1H, dd, *J* = 13.5, 6 Hz), 1.99 (1H, ddd, *J* = 13.5, 6, ~0 Hz), 2.31 (1H, q, *J* = 7 Hz), 4.27 (1H, ddd, *J* = 10, 6, 6 Hz), 4.31 (1H, dd, *J* = 4.5, 4.5 Hz), 4.41 (1H, ddd, *J* = 5.5, 4.5, ~0 Hz), 5.49 (1H, dddd, *J* = 8, 6, 6, 2 Hz), 5.05 (1H, dd, *J* = 6,

6 Hz), 5.33 (1H, dd, *J* = 15, 6 Hz), 5.69 (1H, dd, *J* = 6, 2 Hz), 6.19 (1H, dd, *J* = 15, 11 Hz) and 6.42 (1H, d, *J* = 11 Hz); MS *m/z* (rel. int.): 392, 390 and 388 [M]⁺ (8:16:8), 311 and 309 [M – Br]⁺ (57:57), 273 and 271 [M – C₃H₃Br]⁺ (19:18), 244 and 242 [M – C₃H₂Br – C₂H₅]⁺ (20:20), 231 and 229 [M – C₆H₈Br]⁺ (9:10), 149 (53), 125 (72), 109 (99), 107 (66), 81 (81), 79 (100), 77 (65) and 65 (51); HR-MS *m/z*: 389.9643. Calc. for C₁₅H₁₈O₂⁷⁹Br⁸¹Br: 389.9654.

Hydrogenation of 1. Compound **1** (12 mg) was hydrogenated in EtOAc over PtO₂-catalyst. After removal of the catalyst and the solvent, the residual oil was chromatographed on TLC to give **2** (4 mg); oil; $[\alpha]_D^{21} + 9.02^\circ$ (c 0.35); IR $\nu_{\max}^{\text{CHCl}_3}$ cm^{–1}: 1650, 1610, 1320, 1150, 1125, 1097, 1065, 1035, 963 and 918; ¹H NMR: δ 1.12 (3H, t, *J* = 7 Hz), 2.57 (2H, q, *J* = 7 Hz), 4.03 (1H, m), 4.52 (1H, ddd, *J* = 10, 6, 6 Hz), 4.6–4.8 (2H, m), 5.61 (1H, dd, *J* = 13.5, 6 Hz), 6.29 (1H, dd, *J* = 13.5, 10 Hz) and 6.43 (1H, d, *J* = 10 Hz); MS *m/z* (rel. int.): 316 and 314 [M]⁺ (0.4:0.4), 273 and 271 [M – C₃H₇]⁺ (0.2:0.2), 235 [M – Br]⁺ (1.5), 155 [M – C₆H₈Br]⁺ (92) and 71 (100).

Hydrogenation of 2. Compound **2** (4 mg) was hydrogenated in EtOH over PtO₂. After the usual work-up, the resulting oily substance was purified by TLC to give **3** (2 mg); oil; $[\alpha]_D^{20} + 5.13^\circ$ (c 0.20); IR $\nu_{\max}^{\text{CHCl}_3}$ cm^{–1}: 1130, 1077, 1038 and 910; ¹H NMR: δ 4.0–4.4 (2H, m) and 4.6–4.7 (2H, m); MS *m/z* (rel. int.): 240 [M]⁺ (3), 197 [M – C₃H₇]⁺ (35), 155 [M – C₆H₁₃]⁺ (78) and 71 (100).

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REFERENCES

- Irie, T., Suzuki, M., Kurosawa, E. and Masamune, T. (1970) *Tetrahedron* **26**, 3271.
- Suzuki, M. and Kurosawa, E. (1978) *Tetrahedron Letters* 2503.
- Suzuki, M. and Kurosawa, E. (1979) *Bull. Chem. Soc. Jpn.* **52**, 3352.
- Suzuki, M. and Kurosawa, E. (1981) *Tetrahedron Letters* **22**, 3853.
- Ojika, M., Shizuri, Y. and Yamada, K. (1982) *Phytochemistry* **21**, 2410.
- Kigoshi, H., Shizuri, Y., Niwa, H. and Yamada, K. (1981) *Tetrahedron Letters* **22**, 4729.
- Shizuri, Y., Yamada, A. and Yamada, K. (1984) *Phytochemistry* **23**, 2672.
- Suzuki, T., Koizumi, K., Suzuki, M. and Kurosawa, E. (1983) *Chem. Letters* 1639.
- Kinnel, R., Duggan, A. J., Eisner, T. and Meinwald, J. (1977) *Tetrahedron Letters* 3913.
- Stothers, J. B. (1972) in *Carbon-13 NMR Spectroscopy*, p. 184. Academic Press, New York.
- Tori, K., Komono, T. and Nakagawa, T. (1964) *J. Org. Chem.* **29**, 1136.
- Pretsch, E., Clerc, T., Seible, J. and Simon, W. (1981) in *Tabellen zur Strukturaufklärung organischer Verbindungen*, p. H65. Springer, Berlin.
- Irie, T., Izawa, M. and Kurosawa, E. (1970) *Tetrahedron* **26**, 851.
- Lowe, G. (1965) *J. Chem. Soc. Chem. Commun.* 411.
- Irie, T., Suzuki, T., Yasunari, Y., Kurosawa, E. and Masamune, T. (1969) *Tetrahedron* **25**, 459.
- Suzuki, T., Suzuki, M. and Kurosawa, E. (1975) *Tetrahedron Letters* 3057.

*Previously we reported that the sign of the specific rotation of α-bromocuparene was positive. However, further examination showed that the sign is negative.

17. Sims, J. J., Fenical, W., Wing, R. M. and Radlick, P. (1973) *J. Am. Chem. Soc.* **95**, 972.
18. Faulkner, D. J., Stallard, M. O. and Ireland, C. (1974) *Tetrahedron Letters* 3571.
19. Sims, J. J., Fenical, W., Wing, R. M. and Radlick, P. (1971) *J. Am. Chem. Soc.* **93**, 3774.
20. Sims, J. J., Fenical, W., Wing, R. M. and Radlick, P. (1972) *Tetrahedron Letters* 195.
21. Ireland, C., Stallard, M. O. and Faulkner, D. J. (1976) *J. Org. Chem.* **41**, 2461.